

# Fecal microbiota transplantation: an update on the advances in the treatment of complex diseases

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## ABSTRACT

In addition to its proven effectiveness in the treatment of recurrent *Clostridium difficile* infections (rCDI), fecal microbiota transplantation (FMT) is on the rise to become one of the most studied innovative therapies for the treatment of several complex diseases. This review aims to discuss the many promising applications of FMT in intestinal diseases such as inflammatory bowel diseases (IBD) and extraintestinal diseases such as neuropsychiatric disorders, metabolic syndrome, and autoimmune diseases. For each disease group, pre-clinical data will be presented in addition to clinical trial evidence.

## INTRODUCTION

Gut microbiota is a complex microbial consortium composed of more than 1500 species, distributed in more than 50 different phyla<sup>1</sup>, whose most prevalent are *Bacteroidetes* and *Firmicutes* followed by *Proteobacteria*, *Fusobacteria*, *Tenericutes*, *Actinobacteria*, and *Verrucomicrobia* (90% of the total)<sup>2</sup>. Its composition is however variable and depends on the health/disease state of the host, diet, and environment, alterations in the relative abundance of "good bacteria" is related to several human conditions, such as inflammatory bowel diseases (IBD)<sup>3-11</sup>, obesity and diabetes<sup>12-20</sup>, allergy<sup>21-27</sup>, autoimmune diseases<sup>28-31</sup>, and even neuropsychiatric disorders<sup>32-40</sup>.

Fecal microbiota transplantation (FMT) is a therapeutic approach that aims to normalize the composition of a dysbiotic gut through colonization with stool from a healthy donor<sup>41,42</sup>. So far, FMT has proven effective in treating *Clostridium difficile* infections (CDI) with meta-analysis reporting a 92% clinical resolution rate compared to prolonged anti-microbial therapy with vancomycin<sup>42</sup> with a success rate between 20% and 30%<sup>43-45</sup>. Depending on the source of the material for transplantation, FMT can be divided into heterologous (h-FMT) if it is from a different individual and autologous (a-FMT) if it is the patient's own.

## KEYWORDS

FMT

MICROBIOTA

IBD

NEUROPSYCHIATRIC DISORDERS

METABOLIC SYNDROME

AUTOIMMUNE DISEASES

Implementing a-FMT has been hypothesized to outflank the risk of disease transmission and host-graft reaction<sup>46</sup>. Current guidelines for randomized controlled trials recommend using a-FMT as a placebo because of the assumption that it represents the most inert option compared to h-FMT<sup>47</sup>; however, it is important to highlight how the time of collection is fundamental to the outcome of the intervention, this is because stool samples can have a pro or anti-inflammatory effect<sup>48</sup>. Studies have shown that frozen stool samples are non-inferior to fresh samples in terms of efficacy of the treatment<sup>49</sup>; because of this, stool banking is becoming a valid option to collect healthy samples from the patient during the remission phase and to standardize the process<sup>50</sup>.

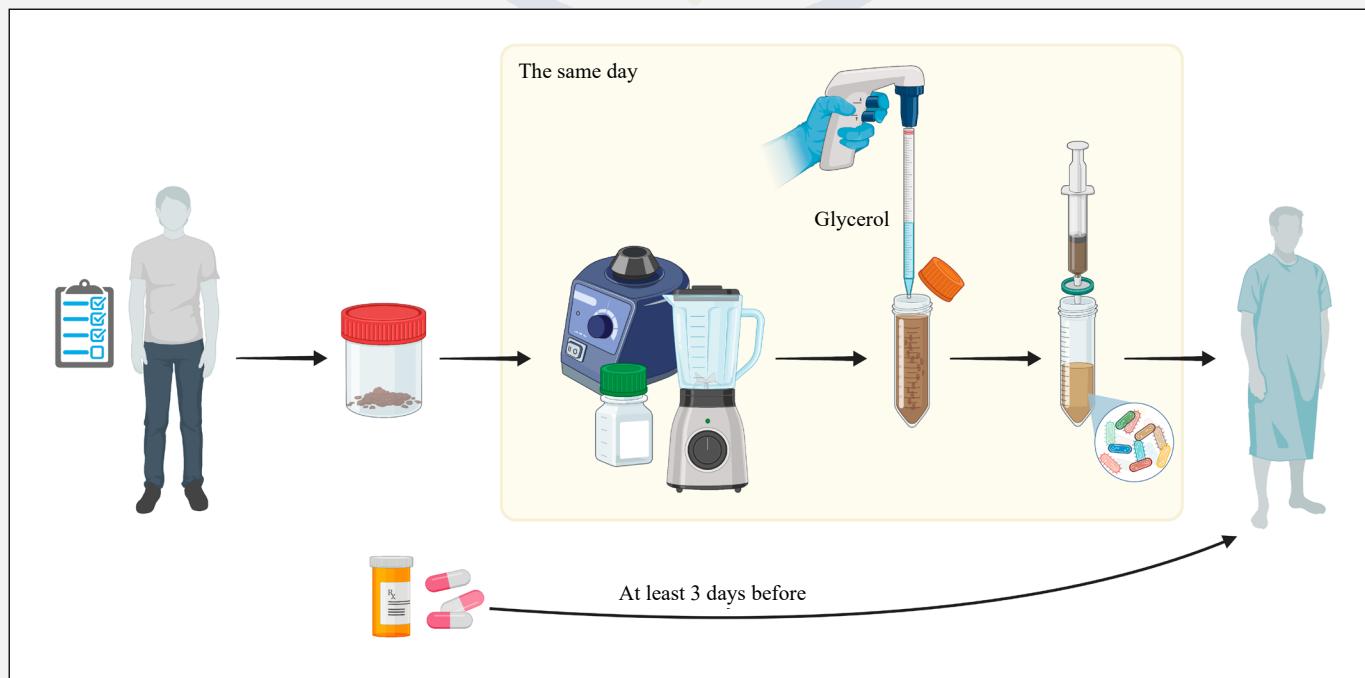
To execute the procedure, stool donors must be tested for viral and non-viral transmittable diseases (e.g., HIV, HBV, *Helicobacter pylori*, *Salmonella*); in addition, a questionnaire is asked to be completed as a screening procedure to exclude unsuitable candidates (e.g., drug use, fat-rich diet, and alcohol use). It was believed that the best stool donor was a close relative, this belief has been disproven<sup>51</sup> since no evidence of any difference in terms of efficacy has been observed; age and gender however must be considered when selecting the donors<sup>52</sup>. Once the quality of the specimen has been evaluated, the fecal material is resuspended in 3 to 5 times the volume of sterile saline solution; in addition, if the sample must be stored for long periods, glycerol should be added up to a final concentration of 10%<sup>53</sup>.

The resuspension in a solvent is fundamental for the homogenization process, which is generally carried out with a mechanical homogenizer or with non-aggressive lysis solutions. Once the sample is dissolved, it must be strained using gauze<sup>49</sup>, tea strainer, or similar device<sup>54</sup> to eliminate bigger pieces that could clog infusion syringes and tubes. The patient treated with FMT must go through a preparation process that consists of a vancomycin treatment at least three days before the infusion, it is important to note that the antibiotic treatment must cease at least 24 to 48 hours before the infusion<sup>49,55-60</sup>. In addition, bowel lavage with polyethylene glycol before the procedure when FMT is performed reduces the abundance of *C. difficile* in the GI tract<sup>49,55,58-62</sup> (Figure 1).

Currently, there is not a defined period of observation after the procedure; however, the European consensus advised conducting at least an 8-week follow-up period after FMT in CDI patients<sup>53,63</sup>. During the follow-up, clinical response is indicated by a reduction of stool frequency and improvement of stool consistency, as well as amelioration of other parameters of disease severity as laboratory parameters, radiological, and/or endoscopic findings<sup>53</sup>.

Although FMT has been utilized more frequently in recent years, many concerns regarding its application, safety, efficacy in the long-term, and the criteria to apply in the selection of the donors are still pending<sup>64</sup>. A case report has observed the transmission of *ESBL Escherichia coli* in two immunocompromised patients

**Figure 1.** Phases of the FMT procedure. The therapy starts with the evaluation of the stool donor. Once it has been thoroughly tested, the sample is blended, enriched with glycerol, and strained. The patient is treated with broad-spectrum antibiotics for at least 3 days prior to the infusion. The figure has been made with biorender.



after FMT from a common stool donor<sup>65</sup>; therefore, a thorough analysis of potential pathogens in the donor's microbiota must be carried out before transplantation. There is also a great need for standardization of the procedure since, so far, every medical center follows its own rules; because of this, the United States and European consensus conference both suggest using a donor questionnaire to meet the exclusion and inclusion criteria<sup>53,63,66-68</sup>.

Since the encouraging results obtained in the treatment of CDI, the range of FMT applications has been constantly expanding<sup>69</sup>. As expected, the first conditions treated with FMT were linked to the GI tract due to the clinical similarities between infectious and non-infectious types of colitis<sup>6,70,71</sup>. Past the gastrointestinal clutter, the applications of FMT rapidly expanded to different areas of extra-gastrointestinal infections in later years, such as obesity and metabolic syndrome<sup>72-77</sup>. Several case reports and animal models have too uncovered the plausible helpful impacts of FMT in patients with severe multiple sclerosis<sup>78,79</sup>, autism<sup>80,81</sup>, multidrug-resistant organisms (MDRO) infections<sup>82-84</sup>, and multiple organ dysfunction in critical patients<sup>85</sup>. Moreover, recent studies illustrated the positive impacts of immunotherapy on melanoma<sup>86-88</sup> with FMT in an animal model and clinical trial.

This review aims to update on the latest applications of FMT in the treatment of inflammatory bowel diseases, neuropsychiatric disorders, metabolic syndrome, and autoimmune diseases through *in vivo* experiments and clinical trials.

## MATERIALS AND METHODS

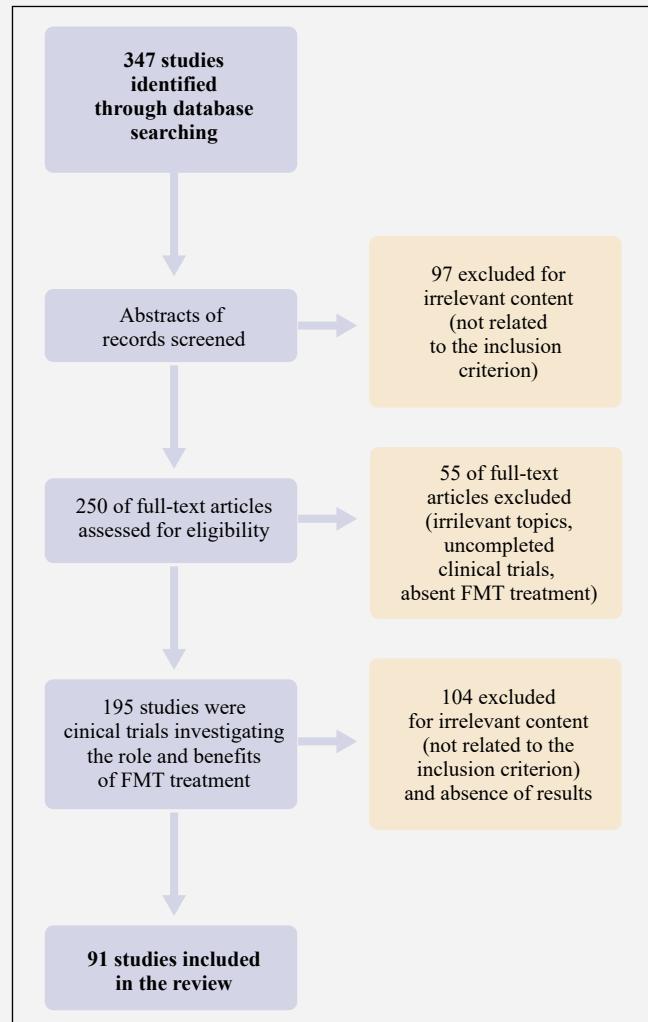
### Search strategy

The literature search was conducted in PubMed (<https://pubmed.ncbi.nlm.nih.gov>) and Scopus (<https://www.scopus.com>) databases considering articles published between 2019 and 2022, using the following search string: ("IBD", "ulcerative colitis", "Crohn's disease") AND ("FMT", "innovative therapy"). The "AND" operator was used to create all possible combinations of selected terms. ClinicalTrials.gov (<https://clinicaltrials.gov>) contains information about medical studies in human volunteers and was used to search for ongoing clinical trials (Figure 2).

### Study selection

The initial screening of documents, using abstracts and titles, was carried out, including only English-language research articles, while articles without full text and

**Figure 2.** Flowchart for review of the studies.



abstract, duplicate studies, conferences, review articles, and editorial reports were excluded.

The selected clinical trials included in this review considered new therapies in the experimental phase II, II, or III used in IBD (UC and CD) patients. In addition, the efficacy and safety of FMT and how it could be a supplement in the treatment of IBD were evaluated.

## RESULTS

### FMT in the treatment of inflammatory bowel diseases

Inflammatory bowel diseases (IBDs), primarily ulcerative colitis (UC), Crohn's disease (CD), and irritable bowel disease (IBS) are multifactorial immune-mediated conditions that affect the lower gastrointestinal tract and have been linked to the onset

of colorectal cancer<sup>89</sup>. IBDs have become, especially in newly industrialized countries that are now exposed to a western diet, the growing diseases of the 21<sup>st</sup> century<sup>90,91</sup>. The exact etiopathology of IBDs is unclear, what is known is that it embroils alterations in the immune response with dysbiosis of the gut microbiota; in addition to that, environmental factors and genetic predisposition can contribute to an early onset of IBD<sup>92</sup>. The main focus of the investigation of the pathogenesis of IBD has always been on imbalances of T cell response on a mucosal level. New evidence suggests that the aberrant inflammatory response in IBD patients is also correlated to adaptive and innate immunity<sup>92</sup>. Previous studies have led to believe that CD was driven by a Th1 response and UC by a Th2 response<sup>93,94</sup>; however, this is not necessarily the case since it has been recently observed how Th17 cells are also involved in the inflammatory response in IBDs<sup>95</sup>. On a microbial level, IBD patients are characterized by a lower abundance of Firmicutes, in particular *Clostridium* clusters IV and XIVa (e.g., genera from the *Ruminococcaceae* and *Lachnospiraceae* family), *Lactobacillus* and *Bifidobacterium*<sup>3,8,96</sup>, and an increase of the abundance of members of the Proteobacteria phylum, such as *Enterobacteriaceae*<sup>97,98</sup>, including *Escherichia coli*<sup>3,99,100</sup>.

FMT was first used to treat UC in 1988<sup>101</sup>, since then, many clinical and preclinical studies have been carried out to verify its success rate and safety. Scientific literature is full of examples of murine models of colitis, such as chemically induced (DSS, TNBS, and oxazolone), genetically induced (*Rag1*<sup>-/-</sup>), and spontaneous models (SAMP1/YitFc); each one differs in terms of reliability and complexity. For example, oral administration of the sulfated polysaccharide DSS to mice via drinking water induces severe colitis characterized by weight loss, bloody diarrhea, ulcer formation, loss of epithelial cells, and infiltrations with neutrophils, resembling some features of flares in human UC<sup>102</sup>. Another example is the SAMP1/YitFc mouse strain, which represents a model of CD-like ileitis that is ideal for investigating the pathogenesis of chronic intestinal inflammation<sup>103</sup>; this is because it develops the ileitis spontaneously due to a single susceptibility locus on chromosome 9 (D9Mit123). Because of the positive results obtained on mice, there was a huge call for clinical trials. A single-center, prospective cohort study conducted by Ianiro et al<sup>104</sup>, which consisted of 8 weeks of follow-up, after the FMT treatment, of 18 IBD patients with a *C. difficile* infection, whose disease activity was assessed through Harvey- Bradshaw Index (HBI  $\geq 4$ )<sup>105</sup> for Crohn's disease and partial Mayo score for ulcerative colitis (Mayo  $\geq 2$ )<sup>106</sup>. The primary outcome of this study was to establish if the patient was negative for *C. difficile* toxin at 8 weeks after the infusion. At

the end of the study, the *C. difficile* toxin was negative in 17 patients (94%), and most of them experienced an improvement in the clinical picture, 10 patients were in clinical remission (59%), and in four patients (24%) the disease was ameliorated, while it did not improve in three patients despite CDI decolonization<sup>104</sup>. Another study conducted by Madsen et al<sup>107</sup> had a primary outcome to evaluate the effect of FMT on abdominal pain, stool frequency, and stool form in IBS patients. The randomized, double-blind, placebo-controlled study included 52 adult patients with moderate-to-severe IBS assigned randomly to treatment with FMT capsules or placebo capsules for 12 days; after that, the patients were followed for a total of 6 months. A statistically significant improvement in stool frequency was found in the FMT group; however, statistically significant differences were found between groups at any time during the study.

#### FMT in the treatment of neuropsychiatric disorders

The intestinal microbiome plays major roles in immune, neuroendocrine, and neural pathways<sup>108</sup>. It has been demonstrated that the gut-brain axis can recruit, through microbiota, a bidirectional communication network to regulate brain function, development, and even behavior<sup>109,110</sup>. Intriguingly, several environmental risk factors for schizophrenia (SCZ), including cesarean section<sup>111,112</sup>, early-life stress<sup>113,114</sup>, maternal malnutrition<sup>115,116</sup>, and maternal immune activation<sup>117,118</sup> have been shown to have a profound impact on the gut microbiome of the newborn.

A preclinical study by Zhu et al<sup>119</sup> used FMT to transfer the microbiota of 11 SCZ patients into C57BL/6J mice. At the end of 3 weeks of infusion, the SCZ mice displayed hyper-kinetic behavior, and an altered tryptophan-kynurenine metabolism was observed. Schizophrenia isn't the only neurologic disorder that has been linked to microbiota alterations, Valeri et al<sup>120</sup> worked with 5xFAD mice treated with antibiotics and FMT of the cecal content of young or old mice to demonstrate how the microbiota could induce Alzheimer's disease (AD) in the transplanted mice. An increase in plaque deposition in PFC and hippocampus DG has been observed in addition to a higher level of LPS binding proteins; these observations have been observed in the very early stages of AD in humans<sup>121</sup>. In conclusion, the authors suggest that a correlation between AD and FMT from an old mice microbiota may exist.

The impact of microbiota transplantation on the progression of neuropsychiatric disorders, such as depression and autism, has been assessed in clinical studies. Lahtinen et al<sup>122</sup> (NCT03561519) revealed in a

randomized controlled trial that IBS patients that have received heterologous FMT had a decrease in both IBS symptoms and depression, based on the IBS-Quality of life questionnaire (IBS-QoL) that was asked to fill in. Autism spectrum disorders (ASD) have long been linked to gastrointestinal distress such as constipation and diarrhea<sup>123-125</sup>; these symptoms seem to be partially due to a dysbiotic microbiota<sup>126</sup> which is unable to process metabolites like 4-ethylphenylsulfate, indolepyruvate, and corticosterone<sup>127,128</sup>. An open-label study published in 2017 (NCT02504554)<sup>129</sup> involved 18 ASD-diagnosed children, between the age of 7 and 16, treated with FMT using a high initial dose followed by daily lower maintenance doses for 7-8 weeks. At the end of the study, significant changes in the gut microbiota composition, GI manifestations, and ASD symptoms were observed:

- the average GSRS score (Gastrointestinal Symptom Rating Scale) dropped 82% from the beginning to the end of the treatment and remained improved at the end of the maintenance phase;
- the ASD-linked behaviors evaluated with PGI-II (Parent Global Impressions) assessment score were significantly improved during treatment without relapse 8 weeks after;
- the high-throughput analysis confirmed the success of the engraftment on a microbial level, illustrating that the transferred microbiota changes the gut environment in a way that is more hospitable to recruiting new commensal bacteria.

### FMT in the treatment of metabolic syndrome

Metabolic syndrome (MetS) is a cluster of conditions such as increased blood pressure, high blood sugar, excess body fat in the waist area, and abnormal cholesterol and/or triglyceride levels; this altogether increases the risk of heart disease, stroke, and type 2 diabetes<sup>130</sup>. Depending on which aspect of metabolic syndrome is studied, many strains of mouse models (also genetically induced) can be used (Table 1). A metabolic syndrome-like phenotype can be reproduced in C57BL/6 mice with the introduction of a high-fat diet<sup>155-157</sup>. In a 2020 preclinical study by Yu et al<sup>158</sup>, db/db mice were used as stool donors to demonstrate that their altered microbiota composition could induce alterations in Hippo signaling in pseudo-germ-free mice. This pathway is recognized as a key regulator of organ size and tissue homeostasis<sup>159</sup>. In another study, the lupus-prone NZB/WF1 strain was given a low-fiber diet to demonstrate how this ultimately leads to accelerated lupus pathology and the associated immune dysregulation. This hypothesis is linked to bacterial metabolism, and especially the generation of SCFA, induced by the consumption of a fiber-rich diet. Their data showed that the altered diet induced an increase in inflamed white adipose tissue in the treated mice, in addition, to intestinal leakage and accelerated disease development have been recorded. These two studies demonstrate how the composition of the gut microbiota can lead to metabolic disorders in prone to the disease mice.

**Table 1.** Mouse models implemented for studying MetS. The table shows some of the most frequently used mouse models, each divided based on the disease in which they are used.

Mouse model	Characteristics	Ref.
<i>Models of obesity and diabetes</i>		
<b>Lep<sup>ob/ob</sup></b>	Early onset of pronounced obese phenotype, hyperphagia and hyperleptinemia followed by hyperinsulinemia, insulin resistance, severe obesity, mild diabetes and fatty liver.	131-138
<b>db/db</b>	More severe hyperglycemia and hyperinsulinemia manifest early (10 <sup>th</sup> day from birth) and progressively aggravate till 3 months. Later on, insulin levels fall accompanied by β-cells atrophy.	139,140
<b>s/s</b>	Knock-in mutation which blocks STAT3 pathway inducing leptin resistance, obesity and diabetes.	141,142
<i>Models of hyperlipidemia</i>		
<b>Ldlr<sup>-/-</sup></b>	Knock-out mutation for LDL receptor protein which leads to two-fold increase in cholesterol, seven- to ninefold increase in intermediate density lipoproteins (IDL) in the bloodstream.	143
<b>ApoE<sup>-/-</sup></b>	Hypercholesterolemia, develop severe atherosclerosis.	144,145
<b>High-fat diet-fed A(y)/a Ldlr<sup>-/-</sup></b>	Severe hyperlipidemia, high levels of VLDL and LDL and insulin resistance.	146,147
<i>Models of hypertension</i>		
<b>NZB/WF1</b>	Autoimmune disease model, especially for systemic lupus erythematosus (SLE), develop insulin resistance and hypertension.	148-150
<b>KKY/a</b>	Develop hyperglycemia, hyperinsulinemia, glucose intolerance and obesity by eight weeks of age	151-154

The implementation of FMT for clinical use has gained much attention in recent years. A 2022 randomized, triple-blind study (NCT03477916) aimed to treat obese patients with metabolic syndrome through microbiota transplantation, in addition to a fiber-rich diet; their results showed that show that a low-fermentable fiber supplementation following oral FMT improved insulin sensitivity from baseline to 6 weeks in the patients. Mounting preclinical and clinical evidence has proved the causal role of gut microbiota on the pathogenesis of primary hypertension. In a randomized, double-blind trial started in 2021 (NCT04406129), 120 patients aged between 18 and 60 with an established diagnosis of Grade 1 hypertension will be divided into two groups and treated either with FMT capsules or placebo; the assigned treatment will be given three times in the outpatient clinics on day 1, day 7, and day 14 after the trial initiation. The patients will be visited once a month for a follow-up period of 3 months. In each visit, the clinician will perform a comprehensive examination to evaluate the efficacy of FMT for hypertension treatment, including physical examination, blood pressure, heart rate, 24-h ABPM, routine blood tests, routine urine tests, biochemical blood tests, urine micro-albuminuria protein, creatinine, electrocardiography, and arterial stiffness assessments (ABIs, PWVs)<sup>160</sup>. Unfortunately, there are no results available because the research group is still recruiting.

### FMT in the treatment of autoimmune diseases

Autoimmune diseases are a group of over one hundred disorders characterized by the dysfunction of the immune system that attacks the body's tissues, causing inflammation and tissue damage. The inflammation is due to the presence of autoreactive lymphocytes or antibodies that causes the death of a specific subpopulation of cells; the consequence is the release of intracellular molecules that, as a vicious cycle, contribute to the stimulation of the immune system<sup>161,162</sup>. The etiopathogenesis of these disorders isn't fully comprehended, but both genetic and environmental factors contribute to the onset<sup>163</sup>. In the last decades, several studies have been conducted on the microbiome's effects on autoimmune patients hypnotizing its role in the loss of immune tolerance and the activation of autoreactive T cells in the intestine. Dysbiosis was associated with the onset or the severity of several autoimmune diseases such as rheumatoid arthritis, spondyloarthritis, Type 1 diabetes, systemic lupus erythematosus, systemic sclerosis, multiple sclerosis, Celiac disease, and Sjögren Syndrome<sup>164-172</sup>. Furthermore, in most of these cases, patients affected

by autoimmune diseases showed, also before the diagnosis, the presence of anti-Saccharomyces cerevisiae autoantibodies, which may modify the microbiome balance<sup>173</sup>.

Consequently, several preclinical studies have been conducted to evaluate the possibility that an FMT can improve the onset and severity of these pathologies. In a multiple sclerosis mice model with experimental autoimmune encephalomyelitis (C57BL/6 mice) was tested the potential efficacy of FMT treatment: mice achieved an amelioration in the dysbiotic gut microbiota, a minor activation of microglia and astrocytes, and protection of the blood-brain barrier, myelin, and axons<sup>174</sup>. In another model of autoimmune disease, the systemic lupus erythematosus mice model (MRL/lpr mice), FMT alleviated the disease severity and progression<sup>175</sup>.

For these reasons, different clinical studies were conducted to test the hypothesis that modifying the microbiome through FMT can improve autoimmune diseases' onset, progress, severity, and treatment also in humans. The randomized controlled trial NTR3697 of FMT on new-onset type 1 diabetes patients indicates that treatment is capable of slowing down the decline in insulin production and preserving beta cell activity<sup>176</sup>. Currently, the following clinical trial is ongoing, exploring the effects of FMT in these patients: NCT04124211, NCT05323162, and NCT04749030 (all in the recruiting phase). In 2022, Huang et al published the results of a 12-week, single-arm pilot clinical trial in which 20 patients with active systemic lupus erythematosus were treated for three weeks with the oral encapsulated fecal microbiome and monitored for 12 weeks: the response rate was 42.12%, there were significant reductions in the severity score and a modification in the bacteria population (a decrease in the inflammation-related bacterial taxa and an increase in SCFAs-producing bacterial taxa)<sup>177</sup>. A case report on FMT performed in a rheumatoid arthritis patient indicates a good potential therapeutical effect<sup>178</sup>. Two clinical trials are ongoing trying to demonstrate this thesis: NCT03944096 (recruiting) and NCT04924270 (not yet recruiting). An exploratory randomized clinical trial on gastroscopic-guided FMT in active peripheral psoriatic arthritis patients (NCT03058900) demonstrated the safety of the technique, but the treatment had no efficacy superior to the placebo<sup>179</sup>. The early phase 1 clinical trial NCT03726645, which started in 2018, will provide new information. The NCT03444220 clinical trial on systemic sclerosis patients demonstrated an amelioration in the lower GI symptoms but demonstrated the presence of adverse events correlated to the gastroduodenoscopy procedure<sup>180</sup>. Is currently ongoing the NCT04300426

clinical trial that will increase the available data. Regarding multiple sclerosis, there are not yet publications available but were started different clinical trials: NCT04150549 (not yet recruiting), NCT04203017 and NCT04096443 (recruiting), NCT03594487 (active, not recruiting), and NCT03975413 (completed). Only one clinical trial (NCT03926286, completed) was performed on Sjogren's Syndrome patients, but the results aren't available. Were also performed clinical trials that included patients with different autoimmune diseases: NCT04924270 (phase 2, not yet recruiting), NCT04014413 (started in 2019, actively recruiting). As these clinical studies will be completed and their results will be published, we will understand better if, as it seems, transplantation can be good therapy for autoimmune diseases. Comparative studies will then be needed to understand which is the best FMT by evaluating both the efficacy and the related adverse effects.

## CONCLUSIONS

It is undeniable that FMT is becoming a well-established therapy with the potential to be used routinely for the treatment of complex diseases. Many preclinical and clinical studies are demonstrating its efficacy in the treatment of extraintestinal conditions such as autism spectrum disorders, type 2 diabetes, and rheumatoid arthritis. The call for standardization of the procedure and the implementation of high-quality randomized controlled trials will give further insights into its long-term effects and benefits. Hence, with the development of personalized medicine, this procedure can be adapted to meet the personal needs of each patient and achieve stable and effective results.

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**CONFLICTS OF INTEREST.** The authors declare that they have no conflicts of interest or competing financial interests.

## REFERENCE

1. Robles-Alonso V, Guarner F. Progreso en el conocimiento de la microbiota intestinal humana [Progress in the knowledge of the intestinal human microbiota]. *Nutr Hosp.* 2013 May-Jun;28(3):553-557. Doi: 10.3305/NH.2013.28.3.6601.
2. Jethwani P, Grover K. Gut Microbiota in Health and Diseases – A Review. *Int J Curr Microbiol* Appl Sci. 2019;8(08):1586-1599. Doi: 10.20546/ijcmas.2019.808.187.
3. Frank DN, St Amand AL, Feldman RA, Boedeker EC, Harpaz N, Pace NR. Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc Natl Acad Sci U S A.* 2007 Aug 21;104(34):13780-13785. Doi: 10.1073/pnas.0706625104.
4. Mentella MC, Scaldaferri F, Pizzoferrato M, Gasbarrini A, Miggiano GAD. Nutrition, IBD and Gut Microbiota: A Review. *Nutrients.* 2020 Mar 29;12(4):944. Doi: 10.3390/nu12040944.
5. Lane ER, Zisman TL, Suskind DL. The microbiota in inflammatory bowel disease: current and therapeutic insights. *J Inflamm Res.* 2017 Jun 10;10:63-73. Doi: 10.2147/JIR.S116088.
6. Hansen JJ, Sartor RB. Therapeutic Manipulation of the Microbiome in IBD: Current Results and Future Approaches. *Curr Treat Options Gastroenterol.* 2015 Mar;13(1):105-120. Doi: 10.1007/S11938-014-0042-7.
7. Frank DN, Robertson CE, Hamm CM, Kpadeh Z, Zhang T, Chen H, Zhu W, Sartor RB, Boedeker EC, Harpaz N, Pace NR, Li E. Disease phenotype and genotype are associated with shifts in intestinal-associated microbiota in inflammatory bowel diseases. *Inflamm Bowel Dis.* 2011 Jan;17(1):179-184. Doi: 10.1002/IBD.21339.
8. Ott SJ, Musfeldt M, Wenderoth DF, Hampe J, Brant O, Fölsch UR, Timmis KN, Schreiber S. Reduction in diversity of the colonic mucosa associated bacterial microflora in patients with active inflammatory bowel disease. *Gut.* 2004 May;53(5):685-693. Doi: 10.1136/GUT.2003.025403.
9. Morgan XC, Tickle TL, Sokol H, Gevers D, Devaney KL, Ward DV, Reyes JA, Shah SA, LeLeiko N, Snapper SB, Bousvaros A, Korzenik J, Sands BE, Xavier RJ, Huttenhower C. Dysfunction of the intestinal microbiome in inflammatory bowel disease and treatment. *Genome Biol.* 2012 Apr 16;13(9):R79. Doi: 10.1186/gb-2012-13-9-r79.
10. Gevers D, Kugathasan S, Denson LA, Vázquez-Baeza Y, Van Treuren W, Ren B, Schwager E, Knights D, Song SJ, Yassour M, Morgan XC, Kostic AD, Luo C, González A, McDonald D, Haberman Y, Walters T, Baker S, Rosh J, Stephens M, Heyman M, Markowitz J, Baldassano R, Griffiths A, Sylvester F, Mack D, Kim S, Crandall W, Hyams J, Huttenhower C, Knight R, Xavier RJ. The treatment-naïve microbiome in new-onset Crohn's disease. *Cell Host Microbe.* 2014 Mar 12;15(3):382-392. Doi: 10.1016/j.chom.2014.02.005.
11. Li J, Butcher J, Mack D, Stintzi A. Functional impacts of the intestinal microbiome in the pathogenesis of inflammatory bowel disease. *Inflamm Bowel Dis.* 2015 Jan;21(1):139-153. Doi: 10.1097/MIB.0000000000000215.

12. Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. *Nat Rev Microbiol.* 2021 Jan;19(1):55-71. Doi: 10.1038/s41579-020-0433-9.
13. Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature.* 2006 Dec 21;444(7122):1027-1031. Doi: 10.1038/nature05414.
14. Tims S, Derom C, Jonkers DM, Vlietinck R, Saris WH, Kleerebezem M, de Vos WM, Zoetendal EG. Microbiota conservation and BMI signatures in adult monozygotic twins. *ISME J.* 2013 Apr;7(4):707-717. Doi: 10.1038/ismej.2012.146.
15. Gophna U, Konikoff T, Nielsen HB. Oscillospira and related bacteria - From metagenomic species to metabolic features. *Environ Microbiol.* 2017 Mar;19(3):835-841. Doi: 10.1111/1462-2920.13658.
16. Liu R, Hong J, Xu X, Feng Q, Zhang D, Gu Y, Shi J, Zhao S, Liu W, Wang X, Xia H, Liu Z, Cui B, Liang P, Xi L, Jin J, Ying X, Wang X, Zhao X, Li W, Jia H, Lan Z, Li F, Wang R, Sun Y, Yang M, Shen Y, Jie Z, Li J, Chen X, Zhong H, Xie H, Zhang Y, Gu W, Deng X, Shen B, Xu X, Yang H, Xu G, Bi Y, Lai S, Wang J, Qi L, Madsen L, Wang J, Ning G, Kristiansen K, Wang W. Gut microbiome and serum metabolome alterations in obesity and after weight-loss intervention. *Nat Med.* 2017 Jul;23(7):859-868. Doi: 10.1038/nm.4358.
17. Allin KH, Tremaroli V, Caesar R, Jensen BAH, Damgaard MTF, Bahl MI, Licht TR, Hansen TH, Nielsen T, Dantoft TM, Linneberg A, Jørgensen T, Vestergaard H, Kristiansen K, Franks PW; IMIDIRECT consortium, Hansen T, Bäckhed F, Pedersen O. Aberrant intestinal microbiota in individuals with prediabetes. *Diabetologia.* 2018 Apr;61(4):810-820. Doi: 10.1007/s00125-018-4550-1.
18. Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, Liang S, Zhang W, Guan Y, Shen D, Peng Y, Zhang D, Jie Z, Wu W, Qin Y, Xue W, Li J, Han L, Lu D, Wu P, Dai Y, Sun X, Li Z, Tang A, Zhong S, Li X, Chen W, Xu R, Wang M, Feng Q, Gong M, Yu J, Zhang Y, Zhang M, Hansen T, Sanchez G, Raes J, Falony G, Okuda S, Almeida M, LeChatelier E, Renault P, Pons N, Batto JM, Zhang Z, Chen H, Yang R, Zheng W, Li S, Yang H, Wang J, Ehrlich SD, Nielsen R, Pedersen O, Kristiansen K, Wang J. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature.* 2012 Oct 4;490(7418):55-60. Doi: 10.1038/nature11450.
19. Zhong H, Ren H, Lu Y, Fang C, Hou G, Yang Z, Chen B, Yang F, Zhao Y, Shi Z, Zhou B, Wu J, Zou H, Zi J, Chen J, Bao X, Hu Y, Gao Y, Zhang J, Xu X, Hou Y, Yang H, Wang J, Liu S, Jia H, Madsen L, Brix S, Kristiansen K, Liu F, Li J. Distinct gut metagenomics and metaproteomics signatures in prediabetics and treatment-naïve type 2 diabetics. *EBioMedicine.* 2019 Sep;47:373-383. Doi: 10.1016/j.ebiom.2019.08.048.
20. Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, Prifti E, Vieira-Silva S, Gudmundsdottir V, Pedersen HK, Arumugam M, Kristiansen K, Voigt AY, Vestergaard H, Hercog R, Costea PI, Kultima JR, Li J, Jørgensen T, Levenez F, Dore J; MetaHIT consortium, Nielsen HB, Brunak S, Raes J, Hansen T, Wang J, Ehrlich SD, Bork P, Pedersen O. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature.* 2015 Dec 10;528(7581):262-266. Doi: 10.1038/nature15766. Epub 2015 Dec 2. Erratum in: *Nature.* 2017 May 3;545(7652):116.
21. Bunyavanich S, Shen N, Grishin A, Wood R, Burks W, Dawson P, Jones SM, Leung DYM, Sampson H, Sicherer S, Clemente JC. Early-life gut microbiome composition and milk allergy resolution. *J Allergy Clin Immunol.* 2016 Oct;138(4):1122-1130. Doi: 10.1016/j.jaci.2016.03.041.
22. Bisgaard H, Li N, Bonnelykke K, Chawes BL, Skov T, Paludan-Müller G, Stokholm J, Smith B, Krogfelt KA. Reduced diversity of the intestinal microbiota during infancy is associated with increased risk of allergic disease at school age. *J Allergy Clin Immunol.* 2011 Sep;128(3):646-52.e1-5. Doi: 10.1016/j.jaci.2011.04.060.
23. McAleer JP, Kolls JK. Contributions of the intestinal microbiome in lung immunity. *Eur J Immunol.* 2018 Jan;48(1):39-49. Doi: 10.1002/eji.201646721.
24. Björkstén B, Sepp E, Julge K, Voor T, Mikelsaar M. Allergy development and the intestinal microflora during the first year of life. *J Allergy Clin Immunol.* 2001 Oct;108(4):516-520. Doi: 10.1067/mai.2001.118130.
25. Woodcock A, Moradi M, Smillie FI, Murray CS, Burnie JP, Custovic A. Clostridium difficile, atopy and wheeze during the first year of life. *Pediatr Allergy Immunol.* 2002 Oct;13(5):357-360. Doi: 10.1034/j.1399-3038.2002.01066.x.
26. Tanaka M, Korenori Y, Washio M, Kobayashi T, Momoda R, Kiyohara C, Kuroda A, Saito Y, Sonomoto K, Nakayama J. Signatures in the gut microbiota of Japanese infants who developed food allergies in early childhood. *FEMS Microbiol Ecol.* 2017 Aug 1;93(8). Doi: 10.1093/femsec/fix099.
27. Roduit C, Frei R, Ferstl R, Loeliger S, Westermann P, Rhyner C, Schiavi E, Barcik W, Rodriguez-Perez N, Wawrzyniak M, Chassard C, Lacroix C, Schmausser-Hechfellner E, Depner M, von Mutius E, Braun-Fahrlander C, Karvonen AM, Kirjavainen PV, Pekkanen J, Dalphin JC, Riedler J, Akdis C, Lauener R, O'Mahony L; PASTURE/EFRAIM study group. High levels of butyrate and propionate in early life are associated with protection against atopy. *Allergy.* 2019 Apr;74(4):799-809. Doi: 10.1111/all.13660.

28. Berer K, Gerdes LA, Cekanaviciute E, Jia X, Xiao L, Xia Z, Liu C, Klotz L, Stauffer U, Baranzini SE, Kümpfel T, Hohlfeld R, Krishnamoorthy G, Wekerle H. Gut microbiota from multiple sclerosis patients enables spontaneous autoimmune encephalomyelitis in mice. *Proc Natl Acad Sci U S A.* 2017 Oct 3;114(40):10719-10724. doi: 10.1073/pnas.1711233114.
29. Xu MQ, Cao HL, Wang WQ, Wang S, Cao XC, Yan F, Wang BM. Fecal microbiota transplantation broadening its application beyond intestinal disorders. *World J Gastroenterol.* 2015 Jan 7;21(1):102-111. doi: 10.3748/wjg.v21.i1.102.
30. Chen F, Stappenbeck TS. Microbiome control of innate reactivity. *Curr Opin Immunol.* 2019 Feb;56:107-113. doi: 10.1016/j.co.2018.12.003.
31. McCoy KD, Ignacio A, Geuking MB. Microbiota and Type 2 immune responses. *Curr Opin Immunol.* 2018 Oct;54:20-27. doi: 10.1016/j.co.2018.05.009.
32. Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, Challis C, Schretter CE, Rocha S, Gradinaru V, Chesselet MF, Keshavarzian A, Shannon KM, Krajmalnik-Brown R, Wittung-Stafshede P, Knight R, Mazmanian SK. Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson's Disease. *Cell.* 2016 Dec 1;167(6):1469-1480.e12. doi: 10.1016/j.cell.2016.11.018.
33. Berer K, Gerdes LA, Cekanaviciute E, Jia X, Xiao L, Xia Z, Liu C, Klotz L, Stauffer U, Baranzini SE, Kümpfel T, Hohlfeld R, Krishnamoorthy G, Wekerle H. Gut microbiota from multiple sclerosis patients enables spontaneous autoimmune encephalomyelitis in mice. *Proc Natl Acad Sci U S A.* 2017 Oct 3;114(40):10719-10724. doi: 10.1073/pnas.1711233114.
34. Cekanaviciute E, Yoo BB, Runia TF, Debelius JW, Singh S, Nelson CA, Kanner R, Bencosme Y, Lee YK, Hauser SL, Crabtree-Hartman E, Sand IK, Gacias M, Zhu Y, Casaccia P, Cree BAC, Knight R, Mazmanian SK, Baranzini SE. Gut bacteria from multiple sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. *Proc Natl Acad Sci U S A.* 2017 Oct 3;114(40):10713-10718. doi: 10.1073/pnas.1711235114. Epub 2017 Sep 11. Erratum in: *Proc Natl Acad Sci U S A.* 2017 Oct 17;114(42):E8943.
35. Scott KA, Ida M, Peterson VL, Prendergast JA, Moloney GM, Izumo T, Murphy K, Murphy A, Ross RP, Stanton C, Dinan TG, Cryan JF. Revisiting Metchnikoff: Age-related alterations in microbiota-gut-brain axis in the mouse. *Brain Behav Immun.* 2017 Oct;65:20-32. doi: 10.1016/j.bbi.2017.02.004.
36. Liu S, Li E, Sun Z, Fu D, Duan G, Jiang M, Yu Y, Mei L, Yang P, Tang Y, Zheng P. Altered gut microbiota and short chain fatty acids in Chinese children with autism spectrum disorder. *Sci Rep.* 2019 Jan 22;9(1):287. doi: 10.1038/s41598-018-36430-z.
37. Luczynski P, Tramullas M, Viola M, Shanahan F, Clarke G, O'Mahony S, Dinan TG, Cryan JF. Microbiota regulates visceral pain in the mouse. *eLife.* 2017 Jun 20;6:e25887. doi: 10.7554/eLife.25887.
38. Friedland RP. Mechanisms of molecular mimicry involving the microbiota in neurodegeneration. *J Alzheimers Dis.* 2015;45(2):349-362. doi: 10.3233/JAD-142841.
39. Itzhaki RF, Lathe R, Balin BJ, Ball MJ, Bearer EL, Braak H, Bullido MJ, Carter C, Clerici M, Cosby SL, Del Tredici K, Field H, Fulop T, Grassi C, Griffin WS, Haas J, Hudson AP, Kamer AR, Kell DB, Licastro F, Letenneur L, Lövheim H, Mancuso R, Miklossy J, Ottó C, Palamara AT, Perry G, Preston C, Pretorius E, Strandberg T, Tabet N, Taylor-Robinson SD, Whittum-Hudson JA. Microbes and Alzheimer's Disease. *J Alzheimers Dis.* 2016;51(4):979-984. doi: 10.3233/JAD-160152.
40. Cowan CSM, Hoban AE, Ventura-Silva AP, Dinan TG, Clarke G, Cryan JF. Gutsy Moves: The Amygdala as a Critical Node in Microbiota to Brain Signaling. *Bioessays.* 2018 Jan;40(1). doi: 10.1002/bies.201700172.
- Gupta A, Khanna S. Fecal Microbiota Transplantation. *JAMA.* 2017 Jul 4;318(1):102. doi: 10.1001/jama.2017.6466.
42. Surawicz CM, Brandt LJ, Binion DG, Ananthakrishnan AN, Curry SR, Gilligan PH, McFarland LV, Mellow M, Zuckerbraun BS. Guidelines for diagnosis, treatment, and prevention of *Clostridium difficile* infections. *Am J Gastroenterol.* 2013 Apr;108(4):478-98; quiz 499. doi: 10.1038/ajg.2013.4.
43. van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, Visser CE, Kuijper EJ, Bartelsman JF, Tijssen JG, Speelman P, Dijkgraaf MG, Keller JJ. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med.* 2013 Jan 31;368(5):407-415. doi: 10.1056/NEJMoa1205037.
44. Gough E, Shaikh H, Manges AR. Systematic review of intestinal microbiota transplantation (fecal bacteriotherapy) for recurrent *Clostridium difficile* infection. *Clin Infect Dis.* 2011 Nov;53(10):994-1002. doi: 10.1093/cid/cir632.
45. Kassam Z, Lee CH, Yuan Y, Hunt RH. Fecal microbiota transplantation for *Clostridium difficile* infection: systematic review and meta-analysis. *Am J Gastroenterol.* 2013 Apr;108(4):500-508. doi: 10.1038/ajg.2013.59.
46. Basson AR, Zhou Y, Seo B, Rodriguez-Palacios A, Cominelli F. Autologous fecal microbiota transplantation for the treatment of inflammatory bowel disease. *Transl Res.* 2020 Dec;226:1-11. doi: 10.1016/j.trsl.2020.05.008

47. Kellermayer R. Fecal microbiota transplantation: great potential with many challenges. *Transl Gastroenterol Hepatol.* 2019 May;25(4):40. Doi: 10.21037/tgh.2019.05.10.
48. Ianiro G, Eusebi LH, Black CJ, Gasbarrini A, Cammarota G, Ford AC. Systematic review with meta-analysis: efficacy of faecal microbiota transplantation for the treatment of irritable bowel syndrome. *Aliment Pharmacol Ther.* 2019 Aug;50(3):240-248. Doi: 10.1111/apt.15330.
49. Lee CH, Steiner T, Petrof EO, Smieja M, Roscoe D, Nematallah A, Weese JS, Collins S, Moayyedi P, Crowther M, Ropeleski MJ, Jayaratne P, Higgins D, Li Y, Rau NV, Kim PT. Frozen vs Fresh Fecal Microbiota Transplantation and Clinical Resolution of Diarrhea in Patients With Recurrent Clostridium difficile Infection: A Randomized Clinical Trial. *JAMA.* 2016 Jan 12;315(2):142-149. Doi: 10.1001/jama.2015.18098.
50. van Lingen E, Terveer EM, van der Meulen-de Jong AE, Vendrik KEW, Verspaget HW, Kuijper EJ, Kassam Z, Keller JJ. Advances in Stool Banking. *Microb Health Dis* 2019;1:e182. Doi: 10.26355/mhd\_20201\_182.
51. Lynch SM, Mu J, Grady JJ, Stevens RG, Devers TJ. Fecal Microbiota Transplantation for Clostridium difficile Infection: A One-Center Experience. *Dig Dis.* 2019;37(6):467-472. Doi: 10.1159/000499873.
52. Vemuri R, Sylvia KE, Klein SL, Forster SC, Plebanski M, Eri R, Flanagan KL. The microgenderome revealed: sex differences in bidirectional interactions between the microbiota, hormones, immunity and disease susceptibility. *Semin Immunopathol.* 2019 Mar;41(2):265-275. Doi: 10.1007/s00281-018-0716-7.
53. Cammarota G, Ianiro G, Tilg H, Rajilić-Stojanović M, Kump P, Satokari R, Sokol H, Arkkila P, Pintus C, Hart A, Segal J, Aloia M, Masucci L, Molinaro A, Scaldaferri F, Gasbarrini G, Lopez-Sanroman A, Link A, de Groot P, de Vos WM, Högenauer C, Malfertheiner P, Mattila E, Milosavljević T, Nieuwdorp M, Sanguinetti M, Simren M, Gasbarrini A; European FMT Working Group. European consensus conference on faecal microbiota transplantation in clinical practice. *Gut.* 2017 Apr;66(4):569-580. Doi: 10.1136/gutjnl-2016-313017.
54. Brandt LJ, Aroniadis OC, Mellow M, Kanatzar A, Kelly C, Park T, Stollman N, Rohlke F, Surawicz C. Long-term follow-up of colonoscopic fecal microbiota transplant for recurrent Clostridium difficile infection. *Am J Gastroenterol.* 2012 Jul;107(7):1079-1087. Doi: 10.1038/ajg.2012.60.
55. van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, Visser CE, Kuijper EJ, Bartelsman JF, Tijssen JG, Speelman P, Dijkgraaf MG, Keller JJ. Duodenal infusion of donor feces for recurrent Clostridium difficile. *N Engl J Med.* 2013 Jan 31;368(5):407-415. Doi: 10.1056/NEJMoa1205037.
56. Cammarota G, Masucci L, Ianiro G, Bibbò S, Dinoi G, Costamagna G, Sanguinetti M, Gasbarrini A. Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent Clostridium difficile infection. *Aliment Pharmacol Ther.* 2015 May;41(9):835-843. Doi: 10.1111/apt.13144.
57. Sokol H, Galperine T, Kapel N, Bourlioux P, Seksik P, Barbut F, Scanzi J, Chast F, Batista R, Joly F, Joly AC, Collignon A, Guery B, Beaugerie L; French Group of Faecal microbiota Transplantation (FGFT). Faecal microbiota transplantation in recurrent Clostridium difficile infection: Recommendations from the French Group of Faecal microbiota Transplantation. *Dig Liver Dis.* 2016 Mar;48(3):242-247. Doi: 10.1016/j.dld.2015.08.017.
58. Kump PK, Krause R, Steininger C, Gröchenig HP, Moschen A, Madl C, Novacek G, Allerberger F, Högenauer C. Empfehlungen zur Anwendung der fäkalen Mikrobiotatransplantation "Stuhltransplantation": Konsensus der Österreichischen Gesellschaft für Gastroenterologie und Hepatologie (ÖGGH) in Zusammenarbeit mit der Österreichischen Gesellschaft für Infektiologie und Tropenmedizin (OEGIT) [Recommendations for the use of faecal microbiota transplantation "stool transplantation": consensus of the Austrian Society of Gastroenterology and Hepatology (ÖGGH) in cooperation with the Austrian Society of Infectious Diseases and Tropical Medicine]. *Z Gastroenterol.* 2014 Dec;52(12):1485-1492. German. Doi: 10.1055/s-0034-1385562.
59. Bakken JS, Borody T, Brandt LJ, Brill JV, Demarco DC, Franzos MA, Kelly C, Khoruts A, Louie T, Martinelli LP, Moore TA, Russell G, Surawicz C; Fecal Microbiota Transplantation Workgroup. Treating Clostridium difficile infection with fecal microbiota transplantation. *Clin Gastroenterol Hepatol.* 2011 Dec;9(12):1044-1049. Doi: 10.1016/j.cgh.2011.08.014.
60. Trubiano JA, Cheng AC, Korman TM, Roder C, Campbell A, May ML, Blyth CC, Ferguson JK, Blackmore TK, Riley TV, Athan E. Australasian Society of Infectious Diseases updated guidelines for the management of Clostridium difficile infection in adults and children in Australia and New Zealand. *Intern Med J.* 2016 Apr;46(4):479-493. Doi: 10.1111/imj.13027.
61. Gorkiewicz G, Thallinger GG, Trajanoski S, Lackner S, Stocker G, Hinterleitner T, Güllý C, Högenauer C. Alterations in the colonic microbiota in response to osmotic diarrhea. *PLoS One.* 2013;8(2):e55817. Doi: 10.1371/journal.pone.0055817.

62. Jalanka J, Salonen A, Salojärvi J, Ritari J, Immonen O, Marciani L, Gowland P, Hoad C, Garsed K, Lam C, Palva A, Spiller RC, de Vos WM. Effects of bowel cleansing on the intestinal microbiota. *Gut*. 2015 Oct;64(10):1562-1568. Doi: 10.1136/gutjnl-2014-307240.
63. Wang JW, Kuo CH, Kuo FC, Wang YK, Hsu WH, Yu FJ, Hu HM, Hsu PI, Wang JY, Wu DC. Fecal microbiota transplantation: Review and update. *J Formos Med Assoc*. 2019 Mar;118 Suppl 1:S23-S31. Doi: 10.1016/j.jfma.2018.08.011.
64. Smith MB, Kelly C, Alm EJ. Policy: How to regulate faecal transplants. *Nature*. 2014 Feb 20;506(7488):290-291. Doi: 10.1038/506290a.
65. DeFilipp Z, Bloom PP, Torres Soto M, Mansour MK, Sater MRA, Huntley MH, Turbett S, Chung RT, Chen YB, Hohmann EL. Drug-Resistant *E. coli* Bacteremia Transmitted by Fecal Microbiota Transplant. *N Engl J Med*. 2019 Nov 21;381(21):2043-2050. Doi: 10.1056/NEJMoa1910437.
66. Smits LP, Bouter KE, de Vos WM, Borody TJ, Nieuwdorp M. Therapeutic potential of fecal microbiota transplantation. *Gastroenterology*. 2013 Nov;145(5):946-53. Doi: 10.1053/j.gastro.2013.08.058.
67. Bakken JS, Borody T, Brandt LJ, Brill JV, Demarco DC, Franzos MA, Kelly C, Khoruts A, Louie T, Martinelli LP, Moore TA, Russell G, Surawicz C; Fecal Microbiota Transplantation Workgroup. Treating *Clostridium difficile* infection with fecal microbiota transplantation. *Clin Gastroenterol Hepatol*. 2011 Dec;9(12):1044-1049. Doi: 10.1016/j.cgh.2011.08.014.
68. Paramsothy S, Borody TJ, Lin E, Finlayson S, Walsh AJ, Samuel D, van den Bogaerde J, Leong RW, Connor S, Ng W, Mitchell HM, Kaakoush N, Kamm MA. Donor Recruitment for Fecal Microbiota Transplantation. *Inflamm Bowel Dis*. 2015 Jul;21(7):1600-1606. Doi: 10.1097/MIB.0000000000000405.
69. Xu MQ, Cao HL, Wang WQ, Wang S, Cao XC, Yan F, Wang BM. Fecal microbiota transplantation broadening its application beyond intestinal disorders. *World J Gastroenterol*. 2015 Jan 7;21(1):102-111. Doi: 10.3748/wjg.v21.i1.102.
70. Zhang W, Zou G, Li B, Du X, Sun Z, Sun Y, Jiang X. Fecal Microbiota Transplantation (FMT) Alleviates Experimental Colitis in Mice by Gut Microbiota Regulation. *J Microbiol Biotechnol*. 2020 Aug 28;30(8):1132-1141. Doi: 10.4014/jmb.2002.02044.
71. Johnsen PH, Hilpusch F, Valle PC, Goll R. The effect of fecal microbiota transplantation on IBS related quality of life and fatigue in moderate to severe non-constipated irritable bowel: Secondary endpoints of a double blind, randomized, placebo-controlled trial. *EBioMedicine*. 2020 Jan;51:102562. Doi: 10.1016/j.ebiom.2019.11.023.
72. Ley RE, Bäckhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut microbial ecology. *Proc Natl Acad Sci U S A*. 2005 Aug 2;102(31):11070-11075. Doi: 10.1073/pnas.0504978102.
73. Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature*. 2006 Dec 21;444(7122):1027-1031. Doi: 10.1038/nature05414.
74. Bäckhed F, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci U S A*. 2007 Jan 16;104(3):979-984. Doi: 10.1073/pnas.0605374104.
75. Vrieze A, Van Nood E, Holleman F, Salojärvi J, Koote RS, Bartelsman JF, Dallinga-Thie GM, Ackermans MT, Serlie MJ, Oozeer R, Derrien M, Druesne A, Van Hylckama Vlieg JE, Bloks VW, Groen AK, Heilig HG, Zoetendal EG, Stroes ES, de Vos WM, Hoekstra JB, Nieuwdorp M. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology*. 2012 Oct;143(4):913-6.e7. Doi: 10.1053/j.gastro.2012.06.031. Epub 2012 Jun 20. Erratum in: *Gastroenterology*. 2013 Jan;144(1):250.
76. Mocanu V, Zhang Z, Deehan EC, Kao DH, Hotte N, Karmali S, Birch DW, Samarasinghe KK, Walter J, Madsen KL. Fecal microbial transplantation and fiber supplementation in patients with severe obesity and metabolic syndrome: a randomized double-blind, placebo-controlled phase 2 trial. *Nat Med*. 2021 Jul;27(7):1272-1279. Doi: 10.1038/s41591-021-01399-2.
77. Yu EW, Gao L, Stastka P, Cheney MC, Mahabamunuge J, Torres Soto M, Ford CB, Bryant JA, Henn MR, Hohmann EL. Fecal microbiota transplantation for the improvement of metabolism in obesity: The FMT-TRIM double-blind placebo-controlled pilot trial. *PLoS Med*. 2020 Mar 9;17(3):e1003051. Doi: 10.1371/journal.pmed.1003051.
78. Li K, Wei S, Hu L, Yin X, Mai Y, Jiang C, Peng X, Cao X, Huang Z, Zhou H, Ma G, Liu Z, Li H, Zhao B. Protection of Fecal Microbiota Transplantation in a Mouse Model of Multiple Sclerosis. *Mediators Inflamm*. 2020 Aug 5;2020:2058272. Doi: 10.1155/2020/2058272.
79. Engen PA, Zaferiou A, Rasmussen H, Naqib A, Green SJ, Fogg LF, Forsyth CB, Raeisi S, Hamaker B, Keshavarzian A. Single-Arm, Non-randomized, Time Series, Single-Subject Study of Fecal Microbiota Transplantation in Multiple Sclerosis. *Front Neurol*. 2020 Sep 8;11:978. Doi: 10.3389/fneur.2020.00978.

80. Kang DW, Adams JB, Coleman DM, Pollard EL, Maldonado J, McDonough-Means S, Caporaso JG, Krajmalnik-Brown R. Long-term benefit of Microbiota Transfer Therapy on autism symptoms and gut microbiota. *Sci Rep.* 2019 Apr 9;9(1):5821. doi: 10.1038/s41598-019-42183-0.
81. Li N, Chen H, Cheng Y, Xu F, Ruan G, Ying S, Tang W, Chen L, Chen M, Lv L, Ping Y, Chen D, Wei Y. Fecal Microbiota Transplantation Relieves Gastrointestinal and Autism Symptoms by Improving the Gut Microbiota in an Open-Label Study. *Front Cell Infect Microbiol.* 2021 Oct 19;11:759435. doi: 10.3389/fcimb.2021.759435. Erratum in: *Front Cell Infect Microbiol.* 2021 Nov 23;11:801376.
82. Seong H, Lee SK, Cheon JH, Yong DE, Koh H, Kang YK, Jeong WY, Lee WJ, Sohn Y, Cho Y, Hyun JH, Baek YJ, Kim MH, Kim JH, Ahn JY, Ku NS, Jeong SJ, Yeom JS, Cho MS, Lee JH, Kim BY, Choi JY. Fecal Microbiota Transplantation for multidrug-resistant organism: Efficacy and Response prediction. *J Infect.* 2020 Nov;81(5):719-725. doi: 10.1016/j.jinf.2020.09.003.
83. Innes AJ, Mullish BH, Ghani R, Szydlo RM, Apperley JF, Olavarria E, Palanicawandar R, Kanfer EJ, Milojkovic D, McDonald JAK, Brannigan ET, Thursz MR, Williams HRT, Davies FJ, Marchesi JR, Pavlù J. Fecal Microbiota Transplant Mitigates Adverse Outcomes Seen in Patients Colonized With Multidrug-Resistant Organisms Undergoing Allogeneic Hematopoietic Cell Transplantation. *Front Cell Infect Microbiol.* 2021 Aug 27;11:684659. doi: 10.3389/fcimb.2021.684659.
84. Bilsen MP, Lambregts MMC, van Prehn J, Kuijper EJ. Faecal microbiota replacement to eradicate antimicrobial resistant bacteria in the intestinal tract – a systematic review. *Curr Opin Gastroenterol.* 2022 Jan 1;38(1):15-25. doi: 10.1097/MOG.0000000000000792.
85. McClave SA, Patel J, Bhutiani N. Should fecal microbial transplantation be used in the ICU? *Curr Opin Crit Care.* 2018 Apr;24(2):105-111. doi: 10.1097/MCC.0000000000000489.
86. Hindson J. FMT for immunotherapy-refractory melanoma. *Nat Rev Gastroenterol Hepatol.* 2021 Feb;18(2):82. doi: 10.1038/s41575-021-00413-9.
87. Davar D, Dzutsev AK, McCulloch JA, Rodrigues RR, Chauvin JM, Morrison RM, Deblasio RN, Menna C, Ding Q, Pagliano O, Zidi B, Zhang S, Badger JH, Vetizou M, Cole AM, Fernandes MR, Prescott S, Costa RGF, Balaji AK, Morgan A, Vujkovic-Cvijin I, Wang H, Borhani AA, Schwartz MB, Dubner HM, Ernst SJ, Rose A, Najjar YG, Belkaid Y, Kirkwood JM, Trinchieri G, Zarour HM. Fecal microbiota transplant overcomes resistance to anti-PD-1 therapy in melanoma patients. *Science.* 2021 Feb 5;371(6529):595-602. doi: 10.1126/science.abf3363.
88. Baruch EN, Youngster I, Ben-Betzalel G, Ortenberg R, Lahat A, Katz L, Adler K, Dick-Necula D, Raskin S, Bloch N, Rotin D, Anafi L, Avivi C, Melnichenko J, Steinberg-Silman Y, Mamtani R, Harati H, Asher N, Shapira-Frommer R, Brosh-Nissimov T, Eshet Y, Ben-Simon S, Ziv O, Khan MAW, Amit M, Ajami NJ, Barshack I, Schachter J, Wargo JA, Koren O, Markel G, Boursi B. Fecal microbiota transplant promotes response in immunotherapy-refractory melanoma patients. *Science.* 2021 Feb 5;371(6529):602-609. doi: 10.1126/science.abb5920.
89. Shaukat A, Virnig DJ, Salfiti NI, Howard DH, Sitaraman SV, Liff JM. Is inflammatory bowel disease an important risk factor among older persons with colorectal cancer in the United States? A population-based case-control study. *Dig Dis Sci.* 2011 Aug;56(8):2378-83. doi: 10.1007/s10620-011-1632-z.
90. Chang M, Chang L, Chang HM, Chang F. Intestinal and Extraintestinal Cancers Associated With Inflammatory Bowel Disease. *Clin Colorectal Cancer.* 2018 Mar;17(1):e29-e37. doi: 10.1016/j.clcc.2017.06.009.
91. Ng SC, Shi HY, Hamidi N, Underwood FE, Tang W, Benchimol EI, Panaccione R, Ghosh S, Wu JCY, Chan FKL, Sung JJY, Kaplan GG. Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet.* 2017 Dec 23;390(10114):2769-2778. doi: 10.1016/S0140-6736(17)32448-0. Erratum in: *Lancet.* 2020 Oct 3;396(10256):e56.
92. Zhang YZ, Li YY. Inflammatory bowel disease: pathogenesis. *World J Gastroenterol.* 2014 Jan 7;20(1):91-99. doi: 10.3748/wjg.v20.i1.91.
93. Cobrin GM, Abreu MT. Defects in mucosal immunity leading to Crohn's disease. *Immunol Rev.* 2005 Aug;206:277-295. doi: 10.1111/j.0105-2896.2005.00293.x.
94. Targan SR, Karp LC. Defects in mucosal immunity leading to ulcerative colitis. *Immunol Rev.* 2005 Aug;206:296-305. doi: 10.1111/j.0105-2896.2005.00286.x.
95. Geremia A, Jewell DP. The IL-23/IL-17 pathway in inflammatory bowel disease. *Expert Rev Gastroenterol Hepatol.* 2012 Apr;6(2):223-237. doi: 10.1586/egh.11.107
96. Kostic AD, Xavier RJ, Gevers D. The microbiome in inflammatory bowel disease: current status and the future ahead. *Gastroenterology.* 2014 May;146(6):1489-1499. doi: 10.1053/j.gastro.2014.02.009.

97. Eksik P, Rigottier-Gois L, Gramet G, Sutren M, Pochart P, Marteau P, Jian R, Doré J. Alterations of the dominant faecal bacterial groups in patients with Crohn's disease of the colon. *Gut*. 2003 Feb;52(2):237-242. Doi: 10.1136/gut.52.2.237.
98. Baumgart M, Dogan B, Rishniw M, Weitzman G, Bosworth B, Yantiss R, Orsi RH, Wiedmann M, McDonough P, Kim SG, Berg D, Schukken Y, Scherl E, Simpson KW. Culture independent analysis of ileal mucosa reveals a selective increase in invasive *Escherichia coli* of novel phylogeny relative to depletion of Clostridiales in Crohn's disease involving the ileum. *ISME J*. 2007 Sep;1(5):403-418. Doi: 10.1038/ismej.2007.52.
99. Sartor RB. Therapeutic correction of bacterial dysbiosis discovered by molecular techniques. *Proc Natl Acad Sci U S A*. 2008 Oct 28;105(43):16413-16414. Doi: 10.1073/pnas.0809363105.
100. Mangin I, Bonnet R, Seksik P, Rigottier-Gois L, Sutren M, Bouhnik Y, Neut C, Collins MD, Colombel JF, Marteau P, Doré J. Molecular inventory of faecal microflora in patients with Crohn's disease. *FEMS Microbiol Ecol*. 2004 Oct 1;50(1):25-36. Doi: 10.1016/j.femsec.2004.05.005.
101. Borody TJ, Campbell J. Fecal microbiota transplantation: current status and future directions. *Expert Rev Gastroenterol Hepatol*. 2011 Dec;5(6):653-655. Doi: 10.1586/egh.11.71.
102. Okayasu I, Hatakeyama S, Yamada M, Ohkusa T, Inagaki Y, Nakaya R. A novel method in the induction of reliable experimental acute and chronic ulcerative colitis in mice. *Gastroenterology*. 1990 Mar;98(3):694-702. Doi: 10.1016/0016-5085(90)90290-h.
103. Pizarro TT, Pastorelli L, Bamias G, Garg RR, Reuter BK, Mercado JR, Chieppa M, Arseneau KO, Ley K, Cominelli F. SAMP1/YitFc mouse strain: a spontaneous model of Crohn's disease-like ileitis. *Inflamm Bowel Dis*. 2011 Dec;17(12):2566-2584. Doi: 10.1002/ibd.21638.
104. Ianiro G, Bibbò S, Porcari S, Settanni CR, Giambò F, Curta AR, Quaranta G, Scaldaferri F, Masucci L, Sanguinetti M, Gasbarrini A, Cammarota G. Fecal microbiota transplantation for recurrent *C. difficile* infection in patients with inflammatory bowel disease: experience of a large-volume European FMT center. *Gut Microbes*. 2021 Jan-Dec;13(1):1994834. Doi: 10.1080/19490976.2021.1994834.
105. Harvey RF, Bradshaw JM. A simple index of Crohn's-disease activity. *Lancet*. 1980 Mar 8;1(8167):514. Doi: 10.1016/s0140-6736(80)92767-1.
106. Lewis JD, Chuai S, Nessel L, Lichtenstein GR, Aberra FN, Ellenberg JH. Use of the noninvasive components of the Mayo score to assess clinical response in ulcerative colitis. *Inflamm Bowel Dis*. 2008 Dec;14(12):1660-1666. Doi: 10.1002/ibd.20520.
107. Madsen AMA, Halkjær SI, Christensen AH, Günther S, Browne PD, Kallemose T, Hansen LH, Petersen AM. The effect of faecal microbiota transplantation on abdominal pain, stool frequency, and stool form in patients with moderate-to-severe irritable bowel syndrome: results from a randomised, double-blind, placebo-controlled study. *Scand J Gastroenterol*. 2021 Jul;56(7):761-769. Doi: 10.1080/00365521.2021.1915375
108. Gacias M, Gaspari S, Santos PM, Tamburini S, Andrade M, Zhang F, Shen N, Tolstikov V, Kiebisch MA, Dupree JL, Zachariou V, Clemente JC, Casaccia P. Microbiota-driven transcriptional changes in prefrontal cortex override genetic differences in social behavior. *eLife*. 2016 Apr 20;5:e13442. Doi: 10.7554/eLife.13442.
109. Dinan TG, Cryan JF. Gut-brain axis in 2016: Brain-gut-microbiota axis – mood, metabolism and behaviour. *Nat Rev Gastroenterol Hepatol*. 2017 Feb;14(2):69-70. Doi: 10.1038/nrgastro.2016.200.
110. Borre YE, O'Keeffe GW, Clarke G, Stanton C, Dinan TG, Cryan JF. Microbiota and neurodevelopmental windows: implications for brain disorders. *Trends Mol Med*. 2014 Sep;20(9):509-518. Doi: 10.1016/j.molmed.2014.05.002.
111. Fond G, Bulzacka E, Boyer L, Llorca PM, Godin O, Brunel L, Andrianarisoa MG, Aouizerate B, Berna F, Capdevielle D, Chereau I, Denizot H, Dorey JM, Dubertret C, Dubreucq J, Faget C, Gabayet F, Le Strat Y, Micoulaud-Franchi JA, Misrahi D, Rey R, Richieri R, Roger M, Passerieux C, Schandrin A, Urbach M, Vidalhet P, Schürhoff F, Leboyer M; FondaMental Academic Centers of Expertise for Schizophrenia (FACE-SZ) Group. Birth by cesarean section and schizophrenia: results from the multicenter FACE-SZ data-set. *Eur Arch Psychiatry Clin Neurosci*. 2017 Sep;267(6):587-594. Doi: 10.1007/s00406-016-0708-3.
112. Moya-Pérez A, Luczynski P, Renes IB, Wang S, Borre Y, Anthony Ryan C, Knol J, Stanton C, Dinan TG, Cryan JF. Intervention strategies for cesarean section-induced alterations in the microbiota-gut-brain axis. *Nutr Rev*. 2017 Apr 1;75(4):225-240. Doi: 10.1093/nutrit/nuw069. <https://doi.org/10.1093/NUTRIT/NUW069>.
113. El Aidy S, Ramsteijn AS, Dini-Andreote F, van Eijk R, Houwing DJ, Salles JF, Olivier JDA. Serotonin Transporter Genotype Modulates the Gut Microbiota Composition in Young Rats, an Effect Augmented by Early Life Stress. *Front Cell Neurosci*. 2017 Aug 3;11:222. Doi: 10.3389/fncel.2017.00222.

114. Scheller-Gilkey G, Thomas SM, Woolwine BJ, Miller AH. Increased early life stress and depressive symptoms in patients with comorbid substance abuse and schizophrenia. *Schizophr Bull.* 2002;28(2):223-231. Doi: 10.1093/oxfordjournals.schbul.a006933.
115. Jašarević E, Rodgers AB, Bale TL. A novel role for maternal stress and microbial transmission in early life programming and neurodevelopment. *Neurobiol Stress.* 2015 Jan 1;1:81-88. Doi: 10.1016/j.ynstr.2014.10.005.
116. St Clair D, Xu M, Wang P, Yu Y, Fang Y, Zhang F, Zheng X, Gu N, Feng G, Sham P, He L. Rates of adult schizophrenia following prenatal exposure to the Chinese famine of 1959-1961. *JAMA.* 2005 Aug 3;294(5):557-562. Doi: 10.1001/jama.294.5.557.
117. Hsiao EY, McBride SW, Hsien S, Sharon G, Hyde ER, McCue T, Codelli JA, Chow J, Reisman SE, Petrosino JF, Patterson PH, Mazmanian SK. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell.* 2013 Dec 19;155(7):1451-1463. Doi: 10.1016/j.cell.2013.11.024.
118. Lipina TV, Zai C, Hlousek D, Roder JC, Wong AH. Maternal immune activation during gestation interacts with Disc1 point mutation to exacerbate schizophrenia-related behaviors in mice. *J Neurosci.* 2013 May 1;33(18):7654-7666. Doi: 10.1523/JNEUROSCI.0091-13.2013.
119. Zhu F, Guo R, Wang W, Ju Y, Wang Q, Ma Q, Sun Q, Fan Y, Xie Y, Yang Z, Jie Z, Zhao B, Xiao L, Yang L, Zhang T, Liu B, Guo L, He X, Chen Y, Chen C, Gao C, Xu X, Yang H, Wang J, Dang Y, Madsen L, Brix S, Kristiansen K, Jia H, Ma X. Transplantation of microbiota from drug-free patients with schizophrenia causes schizophrenia-like abnormal behaviors and dysregulated kynurenone metabolism in mice. *Mol Psychiatry.* 2020 Nov;25(11):2905-2918. Doi: 10.1038/s41380-019-0475-4.
120. Valeri F, Dos Santos Guilherme M, He F, Stoye NM, Schwierz A, Endres K. Impact of the Age of Cecal Material Transfer Donors on Alzheimer's Disease Pathology in 5xFAD Mice. *Microorganisms.* 2021 Dec 9;9(12):2548. Doi: 10.3390/microorganisms9122548.
121. Laske C, Stransky E, Leyhe T, Eschweiler GW, Wittorf A, Richartz E, Bartels M, Buchkremer G, Schott K. Stage-dependent BDNF serum concentrations in Alzheimer's disease. *J Neural Transm (Vienna).* 2006 Sep;113(9):1217-1224. Doi: 10.1007/s00702-005-0397-y.
122. Lahtinen P, Jalanka J, Hartikainen A, Mattila E, Hillilä M, Punkkinen J, Koskenpato J, Anttila VJ, Tillonen J, Satokari R, Arkkila P. Randomised clinical trial: faecal microbiota transplantation versus autologous placebo administered via colonoscopy in irritable bowel syndrome. *Aliment Pharmacol Ther.* 2020 Jun;51(12):1321-1331. Doi: 10.1111/apt.15740.
123. McElhanon BO, McCracken C, Karpen S, Sharp WG. Gastrointestinal symptoms in autism spectrum disorder: a meta-analysis. *Pediatrics.* 2014 May;133(5):872-883. Doi: 10.1542/peds.2013-3995.
124. Adams JB, Johansen LJ, Powell LD, Quig D, Rubin RA. Gastrointestinal flora and gastrointestinal status in children with autism--comparisons to typical children and correlation with autism severity. *BMC Gastroenterol.* 2011 Mar 16;11:22. Doi: 10.1186/1471-230X-11-22.
125. Chaidez V, Hansen RL, Hertz-Pannier I. Gastrointestinal problems in children with autism, developmental delays or typical development. *J Autism Dev Disord.* 2014 May;44(5):1117-1127. Doi: 10.1007/s10803-013-1973-x.
126. Krajmalnik-Brown R, Lozupone C, Kang DW, Adams JB. Gut bacteria in children with autism spectrum disorders: challenges and promise of studying how a complex community influences a complex disease. *Microb Ecol Health Dis.* 2015 Mar 12;26:26914. Doi: 10.3402/mehd.v26.26914.
127. Hsiao EY, McBride SW, Hsien S, Sharon G, Hyde ER, McCue T, Codelli JA, Chow J, Reisman SE, Petrosino JF, Patterson PH, Mazmanian SK. Microbiota modulate behavioral and physiological abnormalities associated with neurodevelopmental disorders. *Cell.* 2013 Dec 19;155(7):1451-1463. Doi: 10.1016/j.cell.2013.11.024.
128. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, Bienenstock J, Cryan JF. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc Natl Acad Sci U S A.* 2011 Sep 20;108(38):16050-16055. Doi: 10.1073/pnas.1102999108.
129. Kang DW, Adams JB, Gregory AC, Borody T, Chittick L, Fasano A, Khoruts A, Geis E, Maldonado J, McDonough-Means S, Pollard EL, Roux S, Sadowsky MJ, Lipson KS, Sullivan MB, Caporaso JG, Krajmalnik-Brown R. Microbiota Transfer Therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: an open-label study. *Microbiome.* 2017 Jan 23;5(1):10. Doi: 10.1186/s40168-016-0225-7.
130. Lear SA, Gasevic D. Ethnicity and Metabolic Syndrome: Implications for Assessment, Management and Prevention. *Nutrients.* 2019 Dec 19;12(1):15. doi: 10.3390/nu12010015.
131. Oler AT, Attie AD. A rapid, microplate SNP genotype assay for the leptinob allele. *J Lipid Res.* 2008 May;49(5):1126-1129. Doi: 10.1194/jlr.D800002-JLR200.
132. Sanna V, Di Giacomo A, La Cava A, Lechner RI, Fontana S, Zappacosta S, Matarese G. Leptin surge precedes onset of autoimmune encephalomyelitis

- and correlates with development of pathogenic T cell responses. *J Clin Invest.* 2003 Jan;111(2):241-250. Doi: 10.1172/JCI16721.
133. Mancuso P, Gottschalk A, Phare SM, Peters-Golden M, Lukacs NW, Huffnagle GB. Leptin-deficient mice exhibit impaired host defense in Gram-negative pneumonia. *J Immunol.* 2002 Apr 15;168(8):4018-4024. Doi: 10.4049/jimmunol.168.8.4018.
134. Siegmund B, Lehr HA, Fantuzzi G. Leptin: a pivotal mediator of intestinal inflammation in mice. *Gastroenterology.* 2002 Jun;122(7):2011-2025. Doi: 10.1053/gast.2002.33631.
135. Malik NM, Carter ND, Murray JF, Scaramuzzi RJ, Wilson CA, Stock MJ. Leptin requirement for conception, implantation, and gestation in the mouse. *Endocrinology.* 2001 Dec;142(12):5198-5202. Doi: 10.1210/endo.142.12.8535.
136. Ewart-Toland A, Mounzih K, Qiu J, Chehab FF. Effect of the genetic background on the reproduction of leptin-deficient obese mice. *Endocrinology.* 1999 Feb;140(2):732-738. Doi: 10.1210/endo.140.2.6470.
137. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature.* 1994 Dec 1;372(6505):425-432. Doi: 10.1038/372425a0. Erratum in: *Nature* 1995 Mar 30;374(6521):479.
138. Coleman DL, Hummel KP. The influence of genetic background on the expression of the obese (Ob) gene in the mouse. *Diabetologia.* 1973 Aug;9(4):287-293. Doi: 10.1007/BF01221856.
139. Hummel KP, Dickie MM, Coleman DL. Diabetes, a new mutation in the mouse. *Science.* 1966 Sep 2;153(3740):1127-1128. Doi: 10.1126/science.153.3740.1127.
140. Coleman DL. Obese and diabetes: two mutant genes causing diabetes-obesity syndromes in mice. *Diabetologia.* 1978 Mar;14(3):141-148. Doi: 10.1007/BF00429772.
141. Bates SH, Stearns WH, Dundon TA, Schubert M, Tso AW, Wang Y, Banks AS, Lavery HJ, Haq AK, Maratos-Flier E, Neel BG, Schwartz MW, Myers MG Jr. STAT3 signalling is required for leptin regulation of energy balance but not reproduction. *Nature.* 2003 Feb 20;421(6925):856-859. Doi: 10.1038/nature01388.
142. Leininger GM, Myers MG Jr. LRb signals act within a distributed network of leptin-responsive neurones to mediate leptin action. *Acta Physiol (Oxf).* 2008 Jan;192(1):49-59. Doi: 10.1111/j.1748-1716.2007.01784.x.
143. Ishibashi S, Brown MS, Goldstein JL, Gerard RD, Hammer RE, Herz J. Hypercholesterolemia in low density lipoprotein receptor knockout mice and its reversal by adenovirus-mediated gene delivery. *J Clin Invest.* 1993 Aug;92(2):883-893. Doi: 10.1172/JCI116663.
144. Plump AS, Smith JD, Hayek T, Aalto-Setälä K, Walsh A, Verstuyft JG, Rubin EM, Breslow JL. Severe hypercholesterolemia and atherosclerosis in apolipoprotein E-deficient mice created by homologous recombination in ES cells. *Cell.* 1992 Oct 16;71(2):343-353. Doi: 10.1016/0092-8674(92)90362-g.
145. Piedrahita JA, Zhang SH, Hagaman JR, Oliver PM, Maeda N. Generation of mice carrying a mutant apolipoprotein E gene inactivated by gene targeting in embryonic stem cells. *Proc Natl Acad Sci USA.* 1992 May 15;89(10):4471-4475. Doi: 10.1073/pnas.89.10.4471.
146. Coenen KR, Gruen ML, Chait A, Hasty AH. Diet-induced increases in adiposity, but not plasma lipids, promote macrophage infiltration into white adipose tissue. *Diabetes.* 2007 Mar;56(3):564-573. Doi: 10.2337/db06-1375.
147. Coenen KR, Hasty AH. Obesity potentiates development of fatty liver and insulin resistance, but not atherosclerosis, in high-fat diet-fed agouti LDLR-deficient mice. *Am J Physiol Endocrinol Metab.* 2007 Aug;293(2):E492-E499. Doi: 10.1152/ajpendo.00171.2007.
148. Ryan MJ, McLemore GR Jr. Hypertension and impaired vascular function in a female mouse model of systemic lupus erythematosus. *Am J Physiol Regul Integr Comp Physiol.* 2007 Feb;292(2):R736-R742. Doi: 10.1152/ajpregu.00168.2006.
149. Ryan MJ, McLemore GR Jr, Hendrix ST. Insulin resistance and obesity in a mouse model of systemic lupus erythematosus. *Hypertension.* 2006 Nov;48(5):988-993. Doi: 10.1161/01.HYP.0000243612.02929.df.
150. Dubois EL, Horowitz RE, Demopoulos HB, Teplitz R. NZB/NZW mice as a model of systemic lupus erythematosus. *JAMA.* 1966 Jan 24;195(4):285-289. Doi: 10.1001/jama.1966.03100040091025.
151. Odaka H, Shino A, Ikeda H, Matsuo T. Antidiobesity and antidiabetic actions of a new potent disaccharidase inhibitor in genetically obese-diabetic mice, KKA(y). *J Nutr Sci Vitaminol (Tokyo).* 1992 Feb;38(1):27-37. Doi: 10.3177/jnsv.38.27.
152. Reddi AS, Camerini-Davalos RA. Hereditary diabetes in the KK mouse: an overview. *Adv Exp Med Biol.* 1988;246:7-15. Doi: 10.1007/978-1-4684-5616-5\_2.
153. Iwatsuka H, Shino A, Suzuki Z. General survey of diabetic features of yellow KK mice. *Endocrinol Jpn.* 1970 Feb;17(1):23-35. Doi: 10.1507/endocrj1954.17.23.
154. Diani AR, Sawada GA, Hannah BA, Jodelis KS, Connell MA, Connell CL, Vidmar TJ, Wyse BM. Analysis of pancreatic islet cells and hormone content in the spontaneously diabetic KKAY mouse by morphometry, immunocytochemistry and radioimmunoassay. *Virchows Arch A Pathol Anat Histopathol.* 1987;412(1):53-61. Doi: 10.1007/BF00750731.

155. Schultz A, Neil D, Aguila MB, Mandarim-de-Lacerda CA. Hepatic adverse effects of fructose consumption independent of overweight/obesity. *Int J Mol Sci.* 2013 Nov 5;14(11):21873-21886. Doi: 10.3390/ijms141121873.
156. Schultz A, Barbosa-da-Silva S, Aguila MB, Mandarim-de-Lacerda CA. Differences and similarities in hepatic lipogenesis, gluconeogenesis and oxidative imbalance in mice fed diets rich in fructose or sucrose. *Food Funct.* 2015 May;6(5):1684-1691. Doi: 10.1039/c5fo00251f.
157. de Oliveira Sá G, Dos Santos Neves V, de Oliveira Fraga SR, Souza-Mello V, Barbosa-da-Silva S. High-intensity interval training has beneficial effects on cardiac remodeling through local renin-angiotensin system modulation in mice fed high-fat or high-fructose diets. *Life Sci.* 2017 Nov 15;189:8-17. Doi: 10.1016/j.lfs.2017.09.012.
158. Yu F, Jiang R, Han W, Zhan G, Xu X, Jiang X, Wang L, Xiang S, Zhou Q, Liu C, Zhu B, Hua F, Yang C. Gut microbiota transplantation from db/db mice induces diabetes-like phenotypes and alterations in Hippo signaling in pseudo germ-free mice. *Aging (Albany NY).* 2020 Nov 20;12(23):24156-24167. Doi: 10.18632/aging.104101.
159. Gumbiner BM, Kim NG. The Hippo-YAP signaling pathway and contact inhibition of growth. *J Cell Sci.* 2014 Feb 15;127(Pt 4):709-717. Doi: 10.1242/jcs.140103.
160. Fan L, Ren J, Chen Y, Wang Y, Guo Z, Bu P, Yang J, Ma W, Zhu B, Zhao Y, Cai J. Effect of fecal microbiota transplantation on primary hypertension and the underlying mechanism of gut microbiome restoration: protocol of a randomized, blinded, placebo-controlled study. *Trials.* 2022 Feb 24;23(1):178. Doi: 10.1186/s13063-022-06086-2.
161. Vangoitsenhoven R, Cresci GAM. Role of Microbiome and Antibiotics in Autoimmune Diseases. *Nutr Clin Pract.* 2020 Jun;35(3):406-416. Doi: 10.1002/ncp.10489.
162. Surace AEA, Hedrich CM. The Role of Epigenetics in Autoimmune/Inflammatory Disease. *Front Immunol.* 2019 Jul 4;10:1525. Doi: 10.3389/fimmu.2019.01525.
163. Rose NR. Prediction and Prevention of Autoimmune Disease in the 21st Century: A Review and Preview. *Am J Epidemiol.* 2016 Mar 1;183(5):403-406. Doi: 10.1093/aje/kwv292.
164. Breban M, Tap J, Leboime A, Said-Nahal R, Langella P, Chiocchia G, Furet JP, Sokol H. Faecal microbiota study reveals specific dysbiosis in spondyloarthritis. *Ann Rheum Dis.* 2017 Sep;76(9):1614-1622. Doi: 10.1136/annrheumdis-2016-211064.
165. Maeda Y, Kurakawa T, Umemoto E, Motooka D, Ito Y, Gotoh K, Hirota K, Matsushita M, Furuta Y, Narazaki M, Sakaguchi N, Kayama H, Nakamura S, Iida T, Saeki Y, Kumanogoh A, Sakaguchi S, Takeda K. Dysbiosis Contributes to Arthritis Development via Activation of Autoreactive T Cells in the Intestine. *Arthritis Rheumatol.* 2016 Nov;68(11):2646-2661. Doi: 10.1002/art.39783.
166. Qi CJ, Zhang Q, Yu M, Xu JP, Zheng J, Wang T, Xiao XH. Imbalance of Fecal Microbiota at Newly Diagnosed Type 1 Diabetes in Chinese Children. *Chin Med J (Engl).* 2016 Jun 5;129(11):1298-1304. Doi: 10.4103/0366-6999.182841.
167. Hevia A, Milani C, López P, Cuervo A, Arboleja S, Duranti S, Turroni F, González S, Suárez A, Gueimonde M, Ventura M, Sánchez B, Margolles A. Intestinal dysbiosis associated with systemic lupus erythematosus. *mBio.* 2014 Sep 30;5(5):e01548-14. Doi: 10.1128/mBio.01548-14.
168. Andréasson K, Alrawi Z, Persson A, Jönsson G, Marsal J. Intestinal dysbiosis is common in systemic sclerosis and associated with gastrointestinal and extraintestinal features of disease. *Arthritis Res Ther.* 2016 Nov 29;18(1):278. Doi: 10.1186/s13075-016-1182-z.
169. Mendez R, Watane A, Farhangi M, Cavuoto KM, Leith T, Budree S, Galor A, Banerjee S. Gut microbial dysbiosis in individuals with Sjögren's syndrome. *Microb Cell Fact.* 2020 Apr 15;19(1):90. Doi: 10.1186/s12934-020-01348-7.
170. Mandl T, Marsal J, Olsson P, Ohlsson B, Andréasson K. Severe intestinal dysbiosis is prevalent in primary Sjögren's syndrome and is associated with systemic disease activity. *Arthritis Res Ther.* 2017 Oct 24;19(1):237. Doi: 10.1186/s13075-017-1446-2.
171. Chen J, Chia N, Kalari KR, Yao JZ, Novotna M, Paz Soldan MM, Luckey DH, Marietta EV, Jeraldo PR, Chen X, Weinshenker BG, Rodriguez M, Kantarci OH, Nelson H, Murray JA, Mangalam AK. Multiple sclerosis patients have a distinct gut microbiota compared to healthy controls. *Sci Rep.* 2016 Jun 27;6:28484. Doi: 10.1038/srep28484.
172. Garcia-Mazcorro JF, Rivera-Gutierrez X, Cobos-Quevedo OJ, Grube-Pagola P, Meixueiro-Daza A, Hernandez-Flores K, Cabrera-Jorge FJ, Vivanco-Cid H, Dowd SE, Remes-Troche JM. First Insights into the Gut Microbiota of Mexican Patients with Celiac Disease and Non-Celiac Gluten Sensitivity. *Nutrients.* 2018 Nov 2;10(11):1641. Doi: 10.3390/nu10111641.
173. Rinaldi M, Perricone R, Blank M, Perricone C, Shoenfeld Y. Anti-Saccharomyces cerevisiae autoantibodies in autoimmune diseases: from bread baking to autoimmunity. *Clin Rev Allergy Immunol.* 2013 Oct;45(2):152-61. Doi: 10.1007/s12016-012-8344-9.

174. Li K, Wei S, Hu L, Yin X, Mai Y, Jiang C, Peng X, Cao X, Huang Z, Zhou H, Ma G, Liu Z, Li H, Zhao B. Protection of Fecal Microbiota Transplantation in a Mouse Model of Multiple Sclerosis. *Mediators Inflamm.* 2020 Aug 5;2020:2058272. Doi: 10.1155/2020/2058272.
175. Zhang Y, Liu Q, Yu Y, Wang M, Wen C, He Z. Early and Short-Term Interventions in the Gut Microbiota Affects Lupus Severity, Progression, and Treatment in MRL/lpr Mice. *Front Microbiol.* 2020 Apr 14;11:628. Doi: 10.3389/fmicb.2020.00628.
176. de Groot P, Nikolic T, Pellegrini S, Sordi V, Imangaliyev S, Rampanelli E, Hanssen N, Attaye I, Bakker G, Duinkerken G, Joosten A, Prodan A, Levin E, Levels H, Potter van Loon B, van Bon A, Brouwer C, van Dam S, Simsek S, van Raalte D, Stam F, Gerdes V, Hoogma R, Diekman M, Gerding M, Rustemeijer C, de Bakker B, Hoekstra J, Zwinderman A, Bergman J, Holleman F, Piemonti L, De Vos W, Roep B, Nieuwdorp M. Faecal microbiota transplantation halts progression of human new-onset type 1 diabetes in a randomised controlled trial. *Gut.* 2021 Jan;70(1):92-105. Doi: 10.1136/gutjnl-2020-322630.
177. Huang C, Yi P, Zhu M, Zhou W, Zhang B, Yi X, Long H, Zhang G, Wu H, Tsokos GC, Zhao M, Lu Q. Erratum to “Safety and efficacy of fecal microbiota transplantation for treatment of systemic lupus erythematosus: An EXPLORER trial” [J. Autoimmun. 130, June 2022, 102844]. *J Autoimmun.* 2022 Jul;131:102862. Doi: 10.1016/j.jaut.2022.102862. Erratum for: *J Autoimmun.* 2022 Jun;130:102844.
178. Zeng J, Peng L, Zheng W, Huang F, Zhang N, Wu D, Yang Y. Fecal microbiota transplantation for rheumatoid arthritis: A case report. *Clin Case Rep.* 2020 Dec 23;9(2):906-909. Doi: 10.1002/ccr3.3677.
179. Kragsnaes MS, Kjeldsen J, Horn HC, Munk HL, Pedersen JK, Just SA, Ahlquist P, Pedersen FM, de Wit M, Möller S, Andersen V, Kristiansen K, Kinggaard Holm D, Holt HM, Christensen R, Ellingsen T. Safety and efficacy of faecal microbiota transplantation for active peripheral psoriatic arthritis: an exploratory randomised placebo-controlled trial. *Ann Rheum Dis.* 2021 Sep;80(9):1158-1167. Doi: 10.1136/annrheumdis-2020-219511.
180. Fretheim H, Chung BK, Didriksen H, Bækkevold ES, Midtvedt Ø, Brønborg C, Holm K, Valeur J, Tennøe AH, Garen T, Midtvedt T, Trøseid M, Zarè H, Lund MB, Hov JR, Lundin KEA, Molberg Ø, Hoffmann-Vold AM. Fecal microbiota transplantation in systemic sclerosis: A double-blind, placebo-controlled randomized pilot trial. *PLoS One.* 2020 May 21;15(5):e0232739. Doi: 10.1371/journal.pone.0232739.